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(54) Title: **MOUSE TCF-3 AND TCF-4 AND TUMOURIGENESIS RELATED TO WNT/WINGLESS SIGNALLING**

(57) Abstract

The invention relates to the field of diagnosis and treatment of cancer and two Wnt/Wingless signalling. The invention provides an isolated and/or recombinant nucleic acid or a specific fragment, homologue or derivative thereof, corresponding to a mammalian Tcf-3 or Tcf-4 gene with a nucleic acid sequence as shown in figure 9 and encoding a Tcf-3 or Tcf-4 protein, or fragment thereof, said protein comprising a β -catenin binding domain and capable of complexing with β -catenin in response to signalling through the Wnt/Wingless cascade. Furthermore, the invention provides a method for identifying a candidate drug comprising use of a cell comprising a nucleic acid provided by the invention.

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MOUSE TCF-3, AND TCF-4 AND TUMOURIGENESIS RELATED TO WNT/WINGLESS SIGNALLING

The invention relates to the field of drug discovery, diagnosis and treatment of cancer and to Wnt/Wingless signalling.

Despite extensive knowledge relating to the
5 multitude of cancer forms (varying in appearance from solid tumours and related metastases in distinct parts of the body to leukaemia's of blood cells that circulate throughout the body, and varying from being totally benign to being aggressively malignant) effective therapy
10 of cancer is difficult and in general restricted to three types; treatment via radiation, via chemotherapy and via surgery. Possibilities for a more specific therapy, directed against the underlying cause of a specific cancer or groups of cancers are currently virtually non-
15 existing. Extensive efforts are directed at providing such specific drugs through drug discovery attempts that try to identify candidate drugs for specific cancer therapy.

Development of cancer often starts with changes in a
20 cell that lead to the unrestricted development and division of that first cell into an ever dividing population of cells. These changes are often an accumulation of mutations or other alterations in key genes that occur chronologically, whereby the mutated
25 cell population loses its original, often specialised character and acquires more and more of a cancerous nature. Normal processes of growth regulation of cells are dysfunctioning in the altered cells. Transcription of genes that are normally only little expressed in said
30 cell type is in cancerous cells no longer controlled.

Activation of transcription of genes by transcription factors that would otherwise be dormant in the specific cell type can for example lead to the so

typical unrestricted growth and neoplastic nature of cancer. An example are mutations in suppressor genes that function normally by generating proteins that are suppressing transcriptional pathways which are no longer
5 of use in a specialised cell. Mutated suppressor genes do not longer help keep the growth of a cell at bay. Drugs directed against or intervening with the specific protein-protein or protein-DNA interactions in transcriptional pathways controlling cell growth or
10 development can be considered typical candidate drugs for later use in specific cancer therapy, especially when such pathways have gone awry and lead to unrestricted growth cells.

The Wnt/Wingless signalling pathway has been most
15 studied in embryonic development of insects or amphibians where it lies at the basis of broad developmental effects. The Wingless signalling pathway establishes segment polarity in *Drosophila*, while the ectopic expression of some members of the Wnt signalling cascade
20 induces the axis duplication in *Xenopus*. A striking symmetry exists between these two pathways (reviewed in 13). Wingless/Wnt signalling is assumed to re-program gene expression in responding cells. Despite the identification of multiple components of this signal
25 transduction pathway, the mechanism by which the re-programming occurs remains a mystery and until very recently, β -catenin represented the most downstream known component in the Wnt signal transduction pathway (reviewed in 23). β -catenin and its *Drosophila* homolog
30 armadillo play dual roles in cell-cell adhesion and in signalling. Both proteins occur in adherent junctions in a complex with α -catenin and cadherin homologs, and both accumulate inside cells in response to Wingless/Wnt signals.

35 The two transcription factors Tcf-1 and Lef-1 are the founding members of a small subfamily of vertebrate HMG box transcription factor genes (5,20,27,28,32). Tcf-1

and Lef-1 were originally defined as lymphoid-specific transcription factors based on their affinity for the enhancers of the CD3 ϵ and the T cell receptor- α genes, respectively. Tcf-1 and Lef-1 were later found to be
5 expressed in largely overlapping, complex patterns during embryogenesis (21). In the course of recent studies, several novel Tcf proteins were cloned from diverse organisms, i.e. dTCF (or Pangolin) from *Drosophila* (4,29) and XTcf-3 from *Xenopus* (16).

10 The biological relevance of the Wnt/Wingless cascade and the interaction between β -catenin and Tcf proteins was tested by ectopic expression of engineered forms of XTcf-3 and Lef-1 in *Xenopus* embryos. Injection of β -catenin into early *Xenopus* embryos induces a secondary
15 axis (8), mimicking the broad effect of Wnt activation. Ectopic expression of a dominant-negative form of XTcf-3, lacking the region required for β -catenin binding, blocks β -catenin's ability to cause axis duplication, and, more importantly, blocks formation of the endogenous axis
20 (16). In concordance with these findings, injection of murine Lef-1 induces the formation of a secondary axis in *Xenopus* embryos (2,10).

It is tempting to speculate that early Wnt-driven developmental signals during embryogenesis in mammals,
25 such as the mouse, are mediated by transcription factors of the Tcf family. However, the available knockout data for mLef-1 and mTcf-1 are not consistent with a role for these transcription factors in Wnt signalling in the mammalian embryo. To the contrary, gene disruption of
30 Tcf-1 yields a severe but restricted T cell developmental defect as the only phenotypic abnormality (31). Mutation of Lef-1 only results in abnormalities in the development of various skin appendages, teeth and the trigeminal nucleus but leaves the immune system and other organ
35 systems intact (30).

A typical example of a transcriptional pathway gone wrong and leading to development of cancer, can be found with adenomatous polyposis coli (APC). Mutations in this gene are the most common disease-causing events in humans, approximately 50% of the population will, during a normal life span, develop colorectal polyps initiated by APC mutations. Individuals who inherit APC mutations develop thousands of colorectal tumours. APC protein interacts with at least six other proteins, β -catenin, γ -catenin, tubulin, EB1, hDLG and ZW3/GSK β kinase that may be involved with communicating APC related growth control. Colon carcinoma cells with mutant APC contain large amounts of monomeric, cytoplasmic β -catenin. Reintroduction of wild-type APC reduces the overall amount of β -catenin.

Recently, a human transcription-factor, hTcf-4, was found to be the only member of the Tcf family expressed in colonic epithelium and in the genetically predisposed colorectal polyps found in APC (14) and it was suggested that wild type APC regulates the interaction between β -catenin with hTcf4 whereas mutant APC leads to accumulation of β -catenin. In colon cells, APC therefore regulates the formation of transcriptionally competent β -catenin/Tcf-4 complexes, and loss of APC function results in uncontrolled activation of Tcf-4 target genes, thereby contributing to colon tumourigenesis. This observation was supported by the finding that the constitutive transcriptional activity of Tcf-4-responsive reporter genes was strongly increased in APC^{-/-} colon carcinoma cells but inactive in non-colonic cells or cell lines of tumour or non-tumour origin. Loss of APC function can thus be seen as a crucial event in the early transformation of colonic epithelium.

Above finding of APC related hTcf-4 deregulation is, albeit enhancing insight in tumourigenesis, restricted to the narrow field of colon carcinoma and has therefor only limited value to the field of diagnosis and treatment of

cancer as a whole. However, there is an obvious need for gaining understanding and access to diagnosis and treatment in a much broader field.

5 The invention provides among others access to and insight in protein-protein or protein-DNA interactions in a transcriptional pathway controlling cell growth or development throughout a wide range of cells and tissues of the body, and provides means, such as nucleic acid,
10 protein, cells and experimental animals and methods to identify candidate drugs, for example for use in cancer therapy.

 The invention provides an isolated and/or recombinant nucleic acid or a specific fragment,
15 homologue or derivative thereof, corresponding to a mammalian Tcf-3 or Tcf-4 gene with a nucleic acid sequence as shown in figure 9 and encoding a Tcf-3 or Tcf-4 protein or fragment thereof, said protein comprising a β -catenin binding domain and capable of
20 complexing with β -catenin in response to signalling through the Wnt/Wingless cascade. Where in this application the definition "nucleic acid" is used, both RNA and DNA, in single or double-stranded fashion, and nucleic acid hybridising thereto is meant. Examples of
25 specific hybridisation are given in the experimental part, wherein among others specific hybridisation with Tcf-3 or Tcf-4 mRNA in various cells or tissues are shown. "Specific fragment" herein meaning a nucleic acid or part thereof that is functionally or structurally
30 related to or hybridising with a distinct nucleic acid or fragment thereof. Typical examples of such a specific fragment as provided by the invention are the β -catenin binding region, the HMG box, and the regions located N-terminally or C-terminally of the HMG box. Comparing
35 mTcf-3 with mTcf-4 shows that said regions for example have 72, 93, 64 and 20% amino acid identity. "Homologue" herein meaning a related nucleic acid that can be found with another species. "Derivative" herein meaning a

nucleic acid that has been derived by genetic modifications, such as deletions, insertions, and mutations from a distinct nucleic acid or fragment thereof. "Corresponding" herein meaning having a nucleic acid sequence homology of at least 80%, more preferably of at least 90%. The invention also provides cDNA, or a specific fragment, homologue or derivative thereof, corresponding to a Tcf-3 or Tcf-4 gene, as for example in the experimental part of the description where cloned cDNAs of these two members of the Tcf family, their expression patterns are determined and their involvement in Wnt signalling is shown.

We have found that Wnt/Wingless signalling regulates the formation of transcriptionally competent β -catenin/Tcf-3 or Tcf-4 complexes, and disturbance of Wnt/Wingless signalling results in uncontrolled activation of Tcf-3 or Tcf-4 target genes, and thus in neoplastic transformation, in a wide variety of cells. The link between Wnt/Wingless signalling and Tcf-3 or Tcf-4 shows that these genes participate in Wnt induced re-programming of gene expression throughout large parts of the organism, also considering the broad patterns of expression seen in the mouse embryo at the various stages of development (see experimental part). Thus, whereas deregulation of APC regulated transcription is limited to hTcf-4 in human colon cells, deregulation of Wnt/Wingless induced transcription of Tcf-3 or Tcf-4 is seen in many cell types of various origin, therewith giving rise to a wide range of neoplasias. Typical examples are foetal cancer, cancers with a stem cell phenotype, testis or ovary teratomas, myeloblastoma, medulloblastoma, mammary or intestinal cancer, and others. Additional cancer types where Wnt/Wingless induced deregulation of transcription of both Tcf-3 or Tcf-4 plays a role can now be identified with a method as provided by the invention, studying Tcf-3 or Tcf-4 transcription activation in cells.

The invention also provides an expression vector comprising a nucleic acid as provided by the invention.

An example of such a vector is given in the experimental part of the description, for example as TCF construct mTcf-3: AA 1-321 or TCF construct mTcf-4: AA 1-331. It is within the skills of the artisan to provide vectors that
5 have been provided with single or multiple nucleic acid changes, deletions and/or insertions or other mutations of a nucleic acid sequence.

The invention also provides a cell comprising a genome in which a nucleic acid sequence corresponding to
10 a nucleic acid according to the invention has been modified. Such a modification can comprise a (for example site-directed or transposon-directed or chemically induced) mutation of a nucleic acid encoding a (fragment) of a Tcf-3 or Tcf-4 gene, be it in the intronic or exonic
15 sequences of said gene. The invention also provides a cell comprising a nucleic acid according to the invention that has been introduced via recombinant means known to the skilled artisan, for example via homologous recombination techniques or by using a vector according
20 to the invention. An example of such a cell according to the invention is provided in the experimental part of the description wherein said cell is a yeast cell, such as for use in a two-hybrid mating assay.

The invention also provides a cell capable of
25 expressing a Tcf-3 or Tcf-4 protein or derivative or fragment thereof, said protein comprising a β -catenin binding domain and capable of complexing with β -catenin in response to signalling through the Wnt/Wingless cascade. Such a cell provided by the invention is for
30 example derived of a cell comprising a genome in which a nucleic acid sequence corresponding to a nucleic acid according to the invention has been modified or derived of a cell comprising a nucleic acid according to the invention. Expression of proteins in recombinant cells is
35 in itself a technique available to the average artisan. An example is provided in the experimental part wherein yeast cells expressing a Tcf-3 or Tcf-4 protein are provided.

The invention also provides a conventional cell capable of expressing Tcf-3 or Tcf-4 mRNA or protein or derivative or fragment thereof, said protein comprising a β -catenin binding domain and capable of complexing with β -catenin in response to signalling through the Wnt/Wingless cascade. Cells expressing a specific mRNA can easily be detected by a skilled artisan, for example by applying hybridisation techniques with nucleic acid probes. Similarly, cells expressing a specific protein can be detected by a skilled artisan by applying techniques such as electrophoreses and/or immunological detection. Such a conventional cell as provided by the invention is for example present in a mouse embryo, a preferred example is a stem cell, but is more preferably derived of a cell line, such as an embryonic stem cell line. Other examples of such a cell or cell line expressing a Tcf-3 or Tcf-4 gene product are given in the experimental part, for instance a PC12 cell, a 3T3 fibroblast, a RBL-5 T-lymphocyte or NS-1 B-lymphocyte are provided by the invention. A preferred embodiment of the invention is a cell derived of a mammary epithial cell line C57MG, which is capable of both a high Tcf-3 or Tcf-4 mRNA and protein expression.

The invention also provides a cell capable of expressing a Tcf-3 or Tcf-4 protein or derivative or fragment thereof, said protein comprising a β -catenin binding domain and capable of complexing with β -catenin in response to signalling through the Wnt/Wingless cascade, wherein said cell is additionally provided with a nucleic acid encoding a repressible Wnt gene or derivative or fragment thereof. At least six Wnt family genes (i.e. Wnt-1, -2, -3a, -4, -5a, -5b) are known, which are expressed in largely overlapping regions within the embryo. Null mutants in at least three Wnt genes have been described. An example of a cell encoding a repressible Wnt gene or derivative or fragment thereof provided by the invention can be found in the

experimental part of this description wherein a 2-69-23 cell (and a method to obtain such a cell) is provided wherein a repressible Wnt-1 gene has been introduced by recombinant techniques. In a cell (for example a 2-69-23 cell) provided by the invention which is additionally comprising a nucleic acid encoding a repressible Wnt gene or derivative or fragment thereof, Wnt induced Tcf-3 or Tcf-4/ β -catenin complexing and subsequent Tcf-target gene trans-activation, and the influence of a variety of compounds thereof, are studied.

The invention also provides a cell capable of expressing a Tcf-3 or Tcf-4 protein or derivative or fragment thereof, said protein comprising a β -catenin binding domain and capable of complexing with β -catenin in response to signalling through the Wnt/Wingless cascade, wherein said cell is additionally provided with a nucleic acid encoding a Tcf-responsive reporter gene. An example of such a cell provided by the invention can be found in the experimental part of this description wherein a 2-69-23 cell is provided wherein a Tcf-reporter gene has been introduced by recombinant techniques. Tcf-responsive reporter (Tcf-reporter) genes are those constructs which comprise a readily detectable or assayable gene (such a luciferase, β -galactosidase, chloramphenicol acetyltransferase) linked in cis to a Tcf-responsive element. Such responsive elements are known in the art and any such elements can be used. An optimal Tcf motif contains the sequence CCTTTGATC. From one to twenty copies, and preferably from three to six copies, of the motif may be used. Mutation of the sequence to CCTTTGGCC abrogates responsiveness. Another necessary part of such constructs is a minimal promotor, such as the c-Fos or the herpesvirus thymidine kinase promotor. Transcription of the reporter gene may be preformed by any means known in the art, usually by assaying for the activity of the encoded gene, although immunological detection methods can also be used. In addition, transcription can be monitored by measuring the

transcribed mRNA directly, typically using oligonucleotide probes. In a cell (for example a 2-69-23 cell) provided by the invention which is additionally comprising a nucleic acid encoding a Tcf-reporter gene, 5 Tcf-target gene activation, and the influence of a variety of compounds thereof, are studied by studying the effect of (modifications of) Tcf or Tcf-complexes and of interactions with compounds therewith, on the transcription of the reporter gene.

10 The invention also provides an animal comprising a genome in which a nucleic acid sequence corresponding to a Tcf nucleic acid according to the invention has been modified or introduced. Such an animal is for example a transgenic animal obtained by modifying an embryonic stem 15 cell. Stem cell modifications, as well as modifications of other cells, are known to the average skilled artisan. Such an animal is for example a homozygous knock-out animal (Tcf-3^{-/-} or Tcf-4^{-/-}) or a heterozygous (Tcf-3^{+/-} of Tcf-4^{+/-}) animal, for example a cross between a knock- 20 out animal with a wild type animal or is otherwise modified in a nucleic acid fragment in its genome that is corresponding to a nucleic acid provided by the invention. A preferred embodiment of the invention is wherein said animal is a mouse.

25 The invention also provides a Tcf-3 or Tcf-4 protein or derivative or fragment thereof, said protein comprising a β -catenin binding domain and capable of complexing with β -catenin in response to signalling through the Wnt/Wingless cascade. Such a protein or 30 derivative or fragment thereof according to the invention is for example encoded by a nucleic acid according to the invention or produced by a cell according to the invention. Proteins, be it natural proteins or recombinant versions thereof can easily be isolated by a 35 skilled artisan, for example when at least a part of the amino acid sequence, or a specific antibody directed against the protein, is provided. The invention provides such an isolated or recombinant Tcf-3 or Tcf-4 protein or

(poly)peptide or derivatives or fragments thereof. In addition, the invention provides an antibody specifically directed against a Tcf-3 or Tcf-4 protein or (poly)peptide or derivative or fragment thereof according to the invention. It is within the skills of the average skilled artisan to provide a (synthetic) antibody directed against a protein or fragment thereof, once the amino acid sequence or the isolated protein is provided. An example of such an antibody, which is specifically directed against hTcf4 and mTcf4 is provided in the experimental part of the description.

The invention also provides a method for identifying a candidate drug comprising use of a cell or an animal or a protein or an antibody according to the invention. Candidate drugs, often first selected or generated via combinatorial chemistry, can now be tested and identified using a method provided by the invention. Such a candidate drug or compound can for example be tested on and selected for their effect on Tcf-3 or Tcf-4 regulated transcription, as for example measured by using a Tcf-responsive reporter gene. For example, a test compound or drug which inhibits the transcription of said reporter gene is a candidate drug for cancer therapy. A preferred embodiment of the invention is a method for identifying a candidate drug wherein said drug is tested on β -catenin/Tcf-3 or β -catenin/Tcf-4 regulated transcription, more preferably when said β -catenin/Tcf-3 or β -catenin/Tcf-4 transcription is part of a Wnt/Wingless signalling cascade. For example, a test compound or drug which inhibits the formation of a β -catenin/Tcf-3 or β -catenin/Tcf-4 complex is a candidate drug for cancer therapy. In a most preferred embodiment of the invention, said testing is performed under conditions wherein Wnt/Wingless signalling can be repressed, thereby for example studying the interaction between Wnt/Wingless

signalling and a test compound. A method provided by the invention for identifying a candidate drug can be accomplished in vitro or in cells. If the method is to be accomplished in cells, then a Tcf-responsive reporter
5 gene must be introduced into the cell. Any means for introducing genetic material into cells can be used, including but not limited to infection, transfection, electroporation. Suitable cells provided by the invention are cells or cell lines recombinantly or conventionally
10 expressing Tcf-3 or Tcf-4 mRNA or Tcf-3 or Tcf-4 protein. A typical example of such a cell suitable for candidate drug testing is a C57MG cell or cell derived thereof, typically provided with a Tcf-responsive reporter gene and/or with a (repressible) Wnt gene. Yet another typical
15 example is an intestinal cell or cell line wherein for example Tcf-4 expression is found.

A preferred embodiment of the invention is a method for identifying a candidate drug wherein said drug is for use in a cancer patient. In the above it is illustrated
20 that deregulation of transcription is an important cause of cancer. Now that it is possible to study deregulation of transcription-activation by Tcf-3 and Tcf-4 in a wide range of cells under many conditions, for example under various conditions of Wnt-signalling, it is possible to
25 test for drugs that intervene in transcription-activation by Tcf-3 and Tcf-4. Such intervening drugs are extremely useful to treat cancers wherein Wnt/Wingless or Tcf-3 or -4 regulated transcription and re-programming gene expression in cells is an underlying cause of the cancer.
30 An example of such a method provided by the invention is wherein said cancer is related to foetal cancer, cancers with a stem cell phenotype, testis or ovary teratomas, myeloblastoma, medulloblastoma, mammary or intestinal cancer, or wherein such a cancer is associated with
35 Wnt/Wingless signalling or defects therein.

The invention also provides a method for diagnosing cancer comprising use of a nucleic acid or a cell or an animal or a protein according or an antibody provided by the invention. For example cells that express Tcf-3 or
5 Tcf-4 gene products or malignant variants thereof can now easily be detected in samples taken from a patient suspected of having a cancer. The invention is described more in detail in the experimental part of the description without limiting the invention thereto.

10

Experimental part

Cloning of mTcf-3 and mTcf-4

15 To isolate Tcf-like genes from the mouse, low-stringency screening of several embryonic cDNA libraries was performed with a mixed probe consisting of the HMG boxes of XTcf-3 (16), chTcf (5) and hTcf-1 (28). All efforts yielded cloned sequences derived from two
20 different Tcf genes. The amino acid sequence directly N-terminal to the HMG boxes were aligned with the corresponding regions encoded by the human genomic fragments of hTCF-3 and hTCF-4 (6). We also isolated full length cDNAs for human Tcf-4, which was 98% identical to
25 mTcf-4 at the amino acid level. An alignment of the amino acid sequences of mTcf-3, hTcf-4, mTcf-4 and XTcf-3 is given in Figure 1. An evolutionary tree was constructed for the available Drosophila and vertebrate Tcf sequences using the program Clustal (Figure 2). mTcf-3 clustered
30 with XTcf-3, while mTcf-4 and hTcf-4 clustered separately. The overall match between mTcf-3 and XTcf-3 is 75%. Both proteins align well in the C-terminal regions downstream of the HMG box (65% identity), the other three mammalian genes contain less conserved C-
35 termini. The mammalian m- or hTcf-3 and XTcf-3 are encoded by orthologous genes.

Two regions of conservation between mTcf-3, mTcf-4 and other members of the family were of particular interest: 1) The N-terminus, which in XTcf-3, hTcf-1 and -4, Lef-1 and dTCF/Pangolin (2,4,10,14,16,29) constitutes the β -catenin interaction domain, and 2) The HMG box DNA-binding domain, which has >90% identity between individual members of the Tcf family. This predicted that mTcf-3 and -4 would bind to Tcf consensus DNA motifs and would interact with β -catenin.

10

Predominantly embryonic expression of mTCF-3 and mTCF-4

We determined the embryonic expression pattern of mTcf-3 and mTcf-4. As depicted in Figure 3A for a longitudinal section of an E6.5 embryo, strong expression of mTcf-3 was observed throughout the embryo proper. In addition, all extra-embryonic tissues with the exception of the ectoplacental cone and the decidua were positive. By E7.5, expression levels were declining, with highest signals remaining in the anterior part of the embryo. The posterior part, including the primitive streak, showed lower expression levels (Figure 3B). At E8.5, the head region still showed expression, especially in the anterior neurectoderm. Expression had almost disappeared in the posterior part of the embryo (Figure 3C). From day 10.5 onwards, mTcf-3 expression became undetectable (not shown).

mTcf-4 expression was undetectable on E6.5, E7.5 and E8.5 (not shown), and was first observed at E10.5 in the roof of the diencephalon and in the anterior part of the mesencephalon, with a sharp posterior boundary (Figure 3D). At all later embryonic stages, high-level expression was mostly restricted to the central nervous system. At E13.5, mTcf-4 mRNA was expressed in the roof of the mesencephalon (the tectum) and in the dorsal thalamus. The expression was sharply demarcated from the

remainder of the thalamus complex by the mTcf-4-negative zona limitans intrathalamica. High level expression was also detected at the di-telencephalic junction (Figure 3E, 3F). At E13.5, relatively low amount of mTcf-4
5 protein was observed in the intestinal epithelium (Figure 3G). The intestinal epithelium was the only embryonic tissue with detectable level of mTcf-4 expression outside of the central nervous system. This expression pattern remained essentially unchanged at E16.5. On
10 coronal sections at E16.5, mTcf-4 mRNA was also detected in the pons cerebri (Figure 3H).

The expression of mTcf-3 and mTcf-4 was also analyzed by Northern blotting on a panel of mouse
15 embryonic (E18.5) and adult tissues, and on several selected cell lines. As expected, no mTcf-3 expression was observed in any of the embryonic or adult tissue RNA samples (not shown). By contrast, high level expression was detected in E14 Embryonic Stem cells, and in the
20 mammary epithelial cell line C57MG, while low level expression occurred in NIH-3T3 fibroblasts (Figure 4B). In line with the in situ data, high level mTcf-4 expression was observed in embryonic brain; all other tissues were essentially negative (Figure 4A), as were
25 all adult tissues tested (not shown). Human Tcf-4 is highly expressed in different cell lines derived from colon adenocarcinomas and hTcf-4 mRNA is clearly detected by in situ analysis in human colon epithelium (14). We noticed a low-level mTcf-4 expression in the intestinal
30 epithelium at E13.5. We realized, that the expression of mTcf-4 in embryonic and adult mouse intestine can not be probably detected by conventional Northern blot using total RNA. We thus performed Northern blot analysis on polyA+ RNA purified from different regions of the mouse
35 intestine. As shown in Figure 4C, mTcf-4 is detected in all parts of the mouse intestine with a clear gradient

along the rostro-caudal axis. The relative values indicate a 10-fold increase of mTcf-4 expression in the distal colon compared to the duodenal part of the small intestine. A similar pattern of the expression was
5 detected for mTcf-3 (Figure 4C). In the tested cell lines, high mTcf-4 expression was observed in the C57MG cell line and in the PC12 pheochromocytoma cell line, Embryonic Stem cells and NIH-3T3 fibroblasts express relatively low level of mTcf-4 mRNA (Figure 4B).

10

mTcf-3 and mTcf-4 interact with β -catenin.

We next wished to obtain molecular support for the involvement of Tcf-3 and Tcf-4 proteins in Wnt/ β -catenin signalling. The interaction between other Tcf members and
15 β -catenin occurs between the N-terminus of the former and the Armadillo repeat of the latter. We utilised the yeast two-hybrid system to demonstrate the interaction. As depicted in Figure 5, two-hybrid interactions were readily detectable between the novel Tcf proteins and β -
20 catenin.

Wnt-1 expression induces nuclear β -catenin/Tcf complexes and activates.

Up till the present invention no experimental proof has linked Wnt stimulation with the induction of β -catenin/Tcf interactions or with induced Tcf reporter gene transcription. The expression of mTcf-3 and mTcf-4 in the Wnt-1-responsive mammary epithelial C57MG cells, allowed us to establish such a direct link. C57MG cells do not express the Wnt-1 gene (19). Upon stimulation by Wnt-1 protein, C57MG cells show morphological transformation and altered growth characteristics (3,12,33). Transient expression of Wnt-1 in C57MG cells results in the increased stability of β -catenin (22). As shown recently, Wnt-1 stimulation reduces ubiquitination and subsequent degradation of β -catenin via the ubiquitin-proteasome pathway (1). In order to study an effect of the Wnt-1 expression on Tcf/ β -catenin mediated transcription we generated C57MG cells carrying the Wnt-1 gene under the control of a tetracycline-repressible promoter. The resulting 2-69-23 cell line shows a strong induction of Wnt-1 mRNA and protein within 24 hours of growth upon removal of tetracycline (Figure 6).

Using a gel retardation/supershift assay, we have previously demonstrated the constitutive presence of nuclear β -catenin/hTcf-4 complex in APC^{-/-} colon carcinoma cells (14) and in colon carcinoma cells with dominant-positive mutations in β -catenin (17). We now applied the same assay to determine whether Wnt stimulation could induce the formation of such complexes. We performed gel retardation with nuclear extracts from 2-69-23 cells stably transfected with a tetracycline-repressible Wnt-1 expression vector. We readily obtained a Tcf/DNA complex from both induced and uninduced cells using an optimal Tcf binding motif as probe. The complex can be supershifted with a monoclonal antibody recognising

human and mouse Tcf-4 (Figure 7). The Wnt-1 induction resulted in the appearance of an additional band, which could be further supershifted with a β -catenin antibody (Figure 7). In the repressed state, a much fainter
5 supershifted band with equal properties was visible in these cells, likely reflecting the low level of leakiness of the repression system (Figure 6). A co-incubation of the binding reaction with both anti β -catenin and anti Tcf-4 antibodies resulted in "super-
10 supershift" confirming simultaneous presence of these proteins in the complex (Figure 7).

To test if Wnt stimulation could activate transcription from Tcf target genes, we performed transient transfections in C57MG and 2-6923 cells with
15 the Tcf reporter plasmid pTOPFLASH or the mutant negative control plasmid pFOPFLASH (Korinek et al., 1997). In the original C57MG cell line, no differential activities between these two reporters were detected (Figure 8). In 2-69-23 cells, Wnt induction resulted in a three-fold
20 stimulation of the Tcf reporter gene transcription as compared to negative control plasmid. Transfection of 2-69-23 cells growing in a presence of tetracycline already showed about 50% difference between the activities of the reporter constructs (Figure 8). This was consistent with
25 the observed formation of a nuclear β -catenin/Tcf complex upon Wnt stimulation. These data provided formal proof that Wnt stimulation indeed induces the formation of nuclear Tcf/ β -catenin complexes and potentially activates transcription from Tcf target genes.

30

The anti Tcf-4 antibody used for the gel retardation/supershift assay does not crossreact with other mouse Tcf proteins.

Discussion

The current study describes the characterisation of two members of the Tcf family. Based on their expression profiles and the physical and functional interactions with the Wnt effector β -catenin, Tcf-3 and Tcf-4 mediate Wnt-driven developmental decisions. Taking advantage of the expression of the Tcf genes in Wnt-1 responsive C57MG cells, we provide direct evidence that Wnt-1 stimulation induces the formation of nuclear complexes between Tcf family members and β -catenin, which in turn can activate transcription from Tcf reporter genes.

Based on sequence comparisons, the mTcf-3 cDNA clone is most closely related to the maternally expressed Xenopus Tcf family member, XTcf-3. XTcf-3 is broadly expressed in the Xenopus embryo (16). Like XTcf-3, mTcf-3 is ubiquitously expressed in the gastrulating embryo. From E7.5, mTcf-3 expression is localised to the anterior part of the embryo and then gradually disappears. An earlier study showed an opposite pattern of Tcf-1 and Lef-1 expression with highest levels of both mRNAs in the posterior part of the embryo. Thus, three Tcf family members are expressed in partially overlapping patterns during early postimplantation development.

Expression patterns of mouse Wnt genes have been studied in great detail. Three Wnt family members, Wnt-3a, Wnt-5a and Wnt-5b, are expressed in discrete spatial domains in the primitive streak. The phenotype of Wnt-3a deficient mice indicates that Wnt-3a regulates dorsal mesoderm fate and is required for generation of all embryonic mesoderm. The highest expression of Tcf-1 and Lef-1 in the mouse embryo at E7.5 is observed in embryonic mesoderm and coincides with Wnt-3a expression. This similarity in expressional profile potentially implies Tcf-1/Lef-1 as downstream mediators of Wnt-3a

signalling in the early stages of mouse embryogenesis. Based on its expression pattern, we believe that mTcf-3 may have a comparable function in early murine embryogenesis, mediating signalling of a, yet to be
5 identified, Wnt factor in the anterior regions of the gastrulating embryo.

During embryogenesis, mTcf-4 displays a highly specific pattern of expression. It appears much later in development than do the other three family members, and
10 its expression is essentially restricted to the di- and mesencephalon, and the intestinal epithelium. Strikingly, the area of mTcf-4 expression in embryonic brain also expresses high levels of Lef-1 (21,30), suggesting a possible redundancy between Tcf-4 and Lef-1 in midbrain
15 development. At least six Wnt family members are expressed in largely overlapping regions within the central nervous system. Expression patterns of the Wnt-1, -3, -3a genes show a temporal and spatial overlap in di- and mesencephalon with that of Tcf-4 and Lef-1 in
20 this region. Therefore, the latter two Tcf genes are likely candidates to mediate signalling of these Wnt factors. Wnt-3 expression displays a clear boundary between the Wnt-3 positive dorsal thalamus and the Wnt-3 negative remainder of diencephalon. The same sharp
25 boundary of expression was observed for Tcf-4 and Lef-1.

Null mutants in three Wnt genes have been described. It is of obvious interest to compare the phenotypes of Wnt knockout mice with those of Tcf/Lef knockout mice.
30 Only the disruption of the Wnt-1 gene yields a defect in midbrain development. The phenotype of the Wnt-1 and Wnt-3a compound mutant mice has demonstrated clear redundancy of these genes in the regulation of dorsal neural tube differentiation. Despite the suggestion of involvement of
35 Tcf factors in Wingless/Wnt signalling (18), the single knockout phenotypes of Tcf-1 and Lef-1 genes have not

implied a role for these two genes in Wnt-1 driven development of the midbrain (15), Wnt-3a regulated somite and tailbud formation, Wnt-4 dependent kidney tubulogenesis or in Wnt-2 regulated development of the placental tissues. The expression patterns of mTcf-3 and mTcf-4 are more restricted in space and time than that of Tcf-1 and Lef-1, and in the case of mTcf-3 implies a specific involvement in early developmental events.

We recently identified hTcf-4 to be expressed in human colonic epithelial cells and cancers derived thereof (14). The available evidence indicates that loss of the tumor-suppressor protein APC or gain-of-function mutations in β -catenin lead to uncontrolled transcriptional activity of Tcf target genes, and -as a consequence- to cellular transformation. The mTcf-4 expression in the embryonic intestinal epithelium suggests a physiological role for this gene in this tissue, possibly in the context of Wnt signalling events that control epithelial homeostasis. In contrast to the brain-specific expression, the intestinal mTcf-4 expression persists to the adult age. mTcf-4 mRNA in mouse intestine is not restricted to the colon, but is also detected in the small intestine. Interestingly, mTcf-4 mRNA shows a gradient with increased levels along the rostro-caudal axis. The same gradient was also detected for mTcf-3.

LEGENDS TO THE FIGURES.

Figure 1. Sequence comparison of mTcf-4, hTcf-4, mTcf-3, and XTcf-3. The highly conserved N-terminal β -catenin interaction domain and the HMG-box region are underlined.

Figure 2. Phylogenetic relationship among members of the Tcf/Lef gene family based on aligned amino acid sequences. Relative branch lengths are drawn proportional

to the number of inferred substitutions per site.

Figure 3. In situ hybridization or immunological analysis of mTcf-3 (A,B,C) and mTcf-4 (D,E,F,G,H) expression on
5 murine embryo sections. mTcf-3 was detected in E6.5 (A), E7.5 (B) and E8.5 (C) embryos. Bright field (left) and the accompanying dark-field (right) pictures of the longitudinal sections are shown. Embryo proper (E), ectoplacental cone (EC), chorion (Ch), neuroectoderm
10 (NE). mTcf-4 expression was first observed at E10.5 (D) in the central nervous system. The expression remains restricted at E13.5 (E). F shows a detail of the head region and G shows a detail of the intestine at E13.5, stained with an antibody directed against Tcf-4. D,E,F
15 and G show parasagittal sections, H shows a coronal section of the head at E16.5. Rhombencephalon (Rh), mesencephalon (Me), diencephalon (Di), telencephalon (Te), tectum (T), dorsal thalamus (DT), di-telencephalic junction (DTJ), (IE) intestinal epithelium, pons (P),
20 esophagus (EP). The horizontal bar in G represents 0.1 mm, in the other sections 0.3 mm.

Figure 4. Northern blot analysis of mTcf-4 and mTcf-3 expression. (A) Tissue specific mTcf-4 expression in
25 mouse embryo at E18.5. The analysis was performed on total RNA isolated from brain (B), gut (G), heart (H), kidney (K), limbs (Lm), liver (Li), lung (Lu), thymus (T). (B) mTcf-4 and mTcf-3 expression in C57MG cells (lane 1), PC12 cells (lane 2), embryonic stem cells (lane
30 3), 3T3 fibroblasts (line 4), RBL-5 T lymphocyte cells (line 5) and NS-1 B lymphocyte cells (line 6). The positions of 18S and 28S ribosomal RNAs are shown. EtBr: ethidium bromide stain. (C) Northern blot analysis of polyA⁺ RNA isolated from different parts of intestine.
35 (1) duodenum, (2) jejunum, (3) ileum, (4) caecum, (5) proximal colon, (6) distal colon. GAPDH: control

hybridization with GAPDH probe.

Figure 5. Two-hybrid mating assay for the interaction of Tcf"s and β -catenin. Baits (hTcf-1 (AA 4-359, AA 176-359 and AA4-63), hLef-1 (AA 1-292), mTcf-3 (AA 1-321), mTcf-4 (AA 1-331), p53) and preys were transformed in MATa or MATa yeast strains respectively, and mated with each other. As shown by growth on selective plates (lower panel) or β -galactosidase assay (not shown), all Tcf"s (but not the N-terminally truncated hTcf-1, or SV40 Large T) interact specifically with β -catenin, and not with a control p53 protein (pVA3). All matings grow on non-selective plates (upper panel).

Figure 6. Inducible expression of Wnt-1 in C57MG cells. (A) Northern blot analysis of Wnt-1 RNA in C57MG cells (clone 2-69-23) carrying a Wnt-1 transgene under the control of a tetracycline (Tet) repressible promoter. PolyA⁺ RNA was isolated from cells cultured in the presence of Tet (50ng/ml) or for 24 hours in the absence of Tet. The Wnt-1 transcript was detected by blot hybridization after gel electrophoresis using a ³²P-labeled probe for Wnt-1. A longer exposure detects the presence of the major Wnt-1 transcript in Tet treated cells. Wnt-1 expression was quantitated with the Computing Densitometer Model 300A from Molecular Dynamics and basal levels were found to be 50 times lower than the de-repressed levels (data not shown). A probe for Neo was used to confirm the presence of equal levels of RNA in each lane (not shown). (B) Western blot analysis of 2-69-23 cells. Equal amounts of Triton X-100 soluble protein from cells cultured in the presence of Tet (50ng/ml) or for 24 hours in the absence of Tet were analysed by immunoblotting with anti-Wnt-1 antibody.

Figure 7. Wnt-1 induces a Tcf/ β -catenin complex in C57MG

cells. Gel retardation assay performed on nuclear extracts from 2-69-23 cells growing in presence or absence (Wnt-1 induction) of Tet (50ng/ml). Samples in lines 1 and 5 were incubated under standard conditions.

5 Anti β -catenin antibody (0.2 mg) was added to the samples in lines 2,6 and 9. A control antibody (human CD4, 0.2 mg) was added to the samples in lines 3 and 7. Anti Tcf-4 antibody (affinity purified, 0.2 mg) was added to samples in lines 8 and 9. Samples in lines 4 and 10 were

10 incubated with a probe mutated in the Tcf binding site.

Figure 8. Wnt-1 activates Tcf reporter construct in C57MG cells.

C57MG cells and 2-69-23 cells growing in the presence or

15 absence of Tet were transfected with 1 mg of the indicated luciferase reporter construct and 1 mg of pCATCONTROL plasmid as internal control. Cells were harvested after 24 hours and luciferase and CAT values were determined.

20

Figure 9. Nucleotide sequences of mTcf-3 (A), mTcf-4 (B) or hTcf-4 (C and D).

METHODS

25

Cloning of Tcf-3 and Tcf-4 gene

Mouse 11-day and 15-day embryo cDNA libraries (Clontech, Palo Alto, USA) or a human 12-week foetal cDNA library were screened at low stringency with a mixed probe

30 derived from HMG boxes of Tcf. Positive clones were subcloned in pBluescriptSK and sequenced.

In situ hybridization.

Dissected embryos were fixed 4 to 18 hours in 4%

35 paraformaldehyde, dehydrated in ethanol and butanol and embedded in paraffin. Embryos were sectioned at 6 mm

thickness and mounted on 3-aminopropyl-triethoxy-silane coated slides. Slides were deparaffined, digested with proteinase K, acetylated with acetic anhydride dissolved in triethanolamine, dehydrated and hybridized overnight
5 at 55°C. Subsequently, high stringency washing was performed in 50% formamide/2 X SSC/10 mM DTT at 65°C. Slides were treated with ribonuclease A, rinsed in 2 X SSC, 0.1 X SDS, dehydrated and dipped in Ilford emulsion G.5. After drying, the slides were exposed for 1 to 10
10 days at 4°C, developed in Kodak D-19 and fixed with Kodak LX-24.

RNA probes

DNA fragments encoding amino acids 270-342 for mTcf-
15 3 and amino acids 258-328 for mTcf-4 were subcloned into pBluescript SK, constructs were linearized and radiolabeled antisense RNA probes were generated from these templates using T7 RNA polymerase and [³⁵S]UTP (Stratagene, La Jolla, CA, USA). Both constructs covered
20 unique DNA sequences. Probes were used for hybridization at 1 x 10⁵ cpm/ml.

RNA analysis

Total RNA from different tissues and cell lines was
25 isolated according to Chomczynski and Sacchi (7). Approximately 15 mg of total RNA was loaded per line and separated on agarose gel containing 2% formaldehyde. The gel was blotted onto a nitrocellulose membrane and probed with ³²P- labelled probes. Probes were the same as
30 for the in situ analysis. PolyA+ RNA was isolated from total RNA using Oligo-dT Dynal beads (Dynal, Oslo, Norway). Approximately. 2 mg. of polyA+ RNA was loaded per line.

35 Tcf-4 antibody

Tcf-4 antibody was raised against the region N-terminal

of the HMG box of Tcf-4 in a mouse. It's specificity was confirmed by screening against COS cells transfected with Tcf-4, Tcf-3, Tcf-1 and Lef-1, and was found positive for human and mouse Tcf-4 only. Staining on mouse and human small intestine shows high levels of Tcf-4 expression in the crypts and villi, with highest levels in the crypts of embryonic small intestine.

Yeast two hybrid analysis

All TCF constructs were created by fusing the regions encoding the indicated amino acids (AA) to the GAL4 binding domain of pMD4. hTcf-1 constructs: AA 4-359, 4-63 and 176-359. hLef-1: AA 1-292. mTcf-3: AA 1-321. mTcf-4: AA 1-331. pVA3 encodes a murine p53/Gal4 DNA-binding domain hybrid in pGBT9 (Clontech). These constructs were transformed in the MATa yeast strain HF7C (Clontech). β -catenin was inserted in frame with the Gal4 activation domain in pGADrx (Stratagene), pTD1 encodes a SV40 large T-antigen in pGAD3F (Clontech), both were transformed into Y187 (MATa) (Clontech). HF7C cells were mated to Y187 cells. The resulting cells were grown on non-selective (leu⁻/trp⁻), or selective (leu⁻/trp⁻/his⁻/25mM 3-Aminotriazol). β -galactosidase filter assays were performed as described in Matchmaker Two Hybrid System (Clontech).

Generation of vectors

The vectors for generating cell lines with a tetracycline repressible Wnt-1 cDNA were kindly provided by H. Bujard (9) and modified as follows: pUHD15-1 was digested with BamHI and EcoRI, blunted, to obtain a 1 kb fragment containing the tetracycline sensitive transactivator (tTA). This fragment was cloned into the StuI site of pHyTCX to generate pHyTCtTA (pHyTCX was generated from pLNCX [GenBank name SYNMMMLPLN3] by John Murphy who replaced the Neomycin resistance gene with the

HygromycinB resistance gene. pUHD10-3 was digested with XhoI and EcoRI to release a 0.46 kb fragment containing a minimal CMV promoter with heptamerized tet-operators. This fragment was cloned into a 6.5 kb EcoRI digested and XhoI partially digested fragment of pROSAbgal (provided by P. Soriano, ref. 25). The resulting vector was named pROTX. An EcoRI fragment containing the Wnt-1 cDNA (from V101) was cloned into the EcoRI site of pROTX to generate pROTWnt-1.

10

Generation of 2-69-23 cell line

C57MG cells carrying the Wnt-1 gene under the control of the tetracycline repressible promoter were generated as follows: C57MG cells were first infected with virus obtained from pROTWnt-1 transfected PE501 cells and selected in G418 until colonies appeared. Clones with a flat morphology were subsequently infected with virus obtained from PE501 cells transfected with pHyTCtTA and selected with HygromycinB in the presence of 5 µg/ml tetracycline. Individual colonies were isolated and screened for their ability to induce Wnt-1 overexpression in the absence of tetracycline.

Wnt-1 Western

Cells were washed in phosphate buffered saline and lysed in ice-cold lysis buffer (1 % Triton X-100, 50 mM Tris-HCl pH8.0, 150 mM NaCl) containing protease inhibitors (1 mM Pefabloc SC [Boehringer Mannheim], 1mM PMSF, 1 ug/ml Leupeptin, 2 ug/ml Aprotinin, 1 ug/ml Pepstatin [Sigma]) and incubated on ice for 20 minutes. Insoluble material was removed by centrifugation at 20,000g for 10 minutes at 4 C. The protein concentration was determined by the method of Bradford (BioRad Protein Assay). Equal amounts of protein were resolved by SDS-polyacrylamide gel electrophoresis and the protein was transferred to nitrocellulose. α-Wnt-1 immunoblots

were blocked in 1 % BSA in TBST and incubated in a 1:1000 dilution of the monoclonal α -Wnt-1 antibody Mcl23 (raised to a Wnt-1 peptide spanning from amino acid 200 to 212 (3)).

5

Cell culture and Wnt-1 induction

C57MG (34) cells were grown in Dulbecco modified Eagle medium (DMEM, Gibco) supplemented with 10% fetal calf serum, 2 mM glutamine, penicillin, streptomycin and 10 mg
10 bovine pancreatic insulin (Sigma) per ml. C57MG cells with a tetracycline-repressible Wnt-1 gene expression (cell line 2-69-23,) were grown in complete medium supplemented with G418 (400 mg/ml, Boehringer), Hygromycin B (100 mg/ml, Boehringer) and tetracycline (50
15 ng/ml, Sigma). For induction studies, cells were washed with PBS, cultivated for additional 24 hours in tetracycline-free medium and then harvested.

Gel retardation assays

20 Assays were performed as described previously (14). Extracts were prepared from intact nuclei of induced and control C57MG cells. Isolated nuclei were washed extensively prior to lysis. The anti β -catenin antibody was purchased from Transduction Laboratories (Lexington, KY). Binding reactions contained 3 mg of nuclear protein,
25 0.5 ng of probe and 100 ng of poly dIdC in 25 ml of binding buffer (60 mM KCl, 1 mM EDTA, 1 mM DTT, 10% glycerol). Samples were incubated 20 minute at room temperature, antibody was added and samples were
30 incubated for another 20-30 minutes, and subjected to non-denaturing polyacrylamide gel electrophoresis.

Cat assays and luciferase assays

2*10⁵ C57MG and 2-69-23 cells (induced or noninduced) were
35 transfected with either pTOPFLASH or pFOPFLASH luciferase reporter constructs (Korinek et al., 1997) using

- LipofectAmine reagent (Gibco). Cells were harvested after 24 hours and lysed in 1 mM DTT, 1 % Triton X-100, 15 % glycerol, 25 mM Tris pH 7.8 and 8 mM $MgCl_2$. Luciferase activity was determined on a Lumac/3M biocounter.
- 5 Plasmid pCATCONTROL (Promega) was used as internal control. CAT values were determined as pristane/xylene-extractable radiolabeled, butyrylated chloramphenicol (van de Wetering et al., 1991).

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CLAIMS

1. An isolated and/or recombinant nucleic acid or a specific fragment, homologue or derivative thereof, corresponding to a mammalian Tcf-3 or Tcf-4 gene with a nucleic acid sequence as shown in figure 9 and encoding a
5 Tcf-3 or Tcf-4 protein, or fragment thereof, said protein comprising a β -catenin binding domain and capable of complexing with β -catenin in response to signalling through the Wnt/Wingless cascade.
2. A nucleic acid according to claim 1 which is a cDNA
10 molecule.
3. A nucleic acid according to claim 1 or 2 which is of mouse origin.
4. An expression vector comprising a nucleic acid according to anyone of claims 1 to 3.
- 15 5. A cell comprising a genome in which a nucleic acid sequence corresponding to a nucleic acid according to anyone of claims 1 to 3 has been modified.
6. A cell comprising a nucleic acid according to anyone of claims 1 to 3 or a vector according to claim 4.
- 20 7. A cell according to claim 6 wherein said cell is a yeast cell.
8. A cell capable of expressing a Tcf-3 or Tcf-4 protein or fragment thereof, said protein comprising a β -catenin binding domain and capable of complexing with β -catenin
25 in response to signalling through the Wnt/Wingless cascade.
9. A cell according to claim 8, said cell derived of a cell according to any of claims 5 to 7.
10. A cell according to claim 8, said cell derived of a
30 cell line.
11. A cell according to claim 10, said cell derived of a mammary epithelial cell line C57MG.
12. A cell according to any of claims 8 to 11, additionally provided with a nucleic acid encoding a
35 repressible Wnt gene or derivative or fragment thereof

and/or provided with a nucleic acid encoding a Tcf-responsive reporter gene.

13. An animal comprising a cell according to claim 5 or 6.
- 5 14. A Tcf-3 or Tcf-4 protein or derivative or fragment thereof, said protein comprising a β -catenin binding domain and capable of complexing with β -catenin in response to signalling through the Wnt/Wingless cascade.
- 15 15. A protein or derivative or fragment thereof according to claim 14 encoded by a nucleic acid according to anyone of claims 1 to 3 or produced by a cell according to anyone of claims 5 to 12.
- 16 16. An antibody specifically directed against a protein or derivative or fragment thereof according to claim 14 or 15.
- 17 17. A method for identifying a candidate drug comprising use of a cell according to any of claims 5 to 12 or an animal according to claim 13 or a protein according to claim 14 or 15 or an antibody according to claim 16.
- 20 18. A method according to claim 17 wherein said drug is for use in a cancer patient.
- 25 19. A method according to claim 18 wherein said cancer is related to foetal cancer, cancers with a stem cell phenotype, testis or ovary teratomas, myeloblastoma, medulloblastoma, mammary or intestinal cancer.
- 30 20. A method for diagnosing cancer comprising use of a nucleic acid according to anyone of claims 1 to 3, a cell according to any of claims 5 to 12 or an animal according to claim 13 or a protein according to claim 14 or 15 or an antibody according to claim 16.

MTCF-4	MPQLNGG-----GGDDLGADELISFKDEG-EQEEKNS	32
HTCF-4	MPQLNGG-----GGDDLGADELISFKDEG-EQEEKNS	32
MTCF-3	MPQLGGRRGGAGGGGGGAGATSGGDDLGADELIPFQDEGGEEQEPSS	50
XTCF-3	MPQLNSG-----GGDELGADELIRFKDEG-EQEEKSP	32
MTCF-4	ENSSAERDLADVKSLSVNESETNQDSSDSEAERRPPRSESEFRDKSRES	82
HTCF-4	ENSSAERDLADVKSLSVNESETNQDSSDSEAERRPPRSESEFRDKSRES	82
MTCF-3	DTASAQRDLDEVKSSLSVNESE-NQSSSDSEAERRPQPARDAF-QKPRDY	98
XTCF-3	<u>GEGSAEGDLADVKSLSVNESE-NHSSSDSEVERRP-PPREAF-EKHRDY</u>	79
MTCF-4	LEEAAGR-QDGGLEFKGPPYPGYPFIMIPDLTSPYLPNGSLSPT-ARTYLQ	130
HTCF-4	LEEAAGR-QDGGLEFKGPPYPGYPFIMIPDLTSPYLPNGSLSPT-ARTYLQ	130
MTCF-3	FAE-VRRPQDGAFFKGGAYPGYPFLMIPDLSSPYLSNGPLSPGGARTYLQ	147
XTCF-3	LTEALRRQQDAAFFKGGPPYAGYPFLMIPDLGGHYLPNGALSPS-ARTYLQ	128
MTCF-4	MKWPLLDVQAGSLQSRQTLKDARSPSPAHI VSNKVPVQHPHHVHPLTPL	180
HTCF-4	MKWPLLDVQAGSLQSRQALKDARSPSPAHI VSNKVPVQHPHHVHPLTPL	180
MTCF-3	MKWPLLDVPSSA-----TVKDTRSPSPAHL-SNKVPVQHPHHMHLTPL	191
XTCF-3	MKWPLLDSPSTA-----GLKDARSPSPAHL-SNKVPVQHPHHMHLTPL	172
MTCF-4	ITYSNEHFTPGNPPPHLPADVDPKTGIPRPPHPPDISPYPLSPGTVGQI	230
HTCF-4	ITYSNEHFTPGNPPPHLPADVDPKTGIPRPPHPPDISPYPLSPGTVGQI	230
MTCF-3	ITYSNDHFSASPPTHLSPEIDPKTGIPRPPHPPSELSPYPLSPGAVGQI	241
XTCF-3	ITYSNEHFSPGTPPGHLSPEIDPKTGIPRPPHPPSELSPYPLSPGAVGQI	222

Fig. 1

Pages 2/18 - 4/18

MISSING UPON TIME OF PUBLICATION

5/18

Fig. 3a

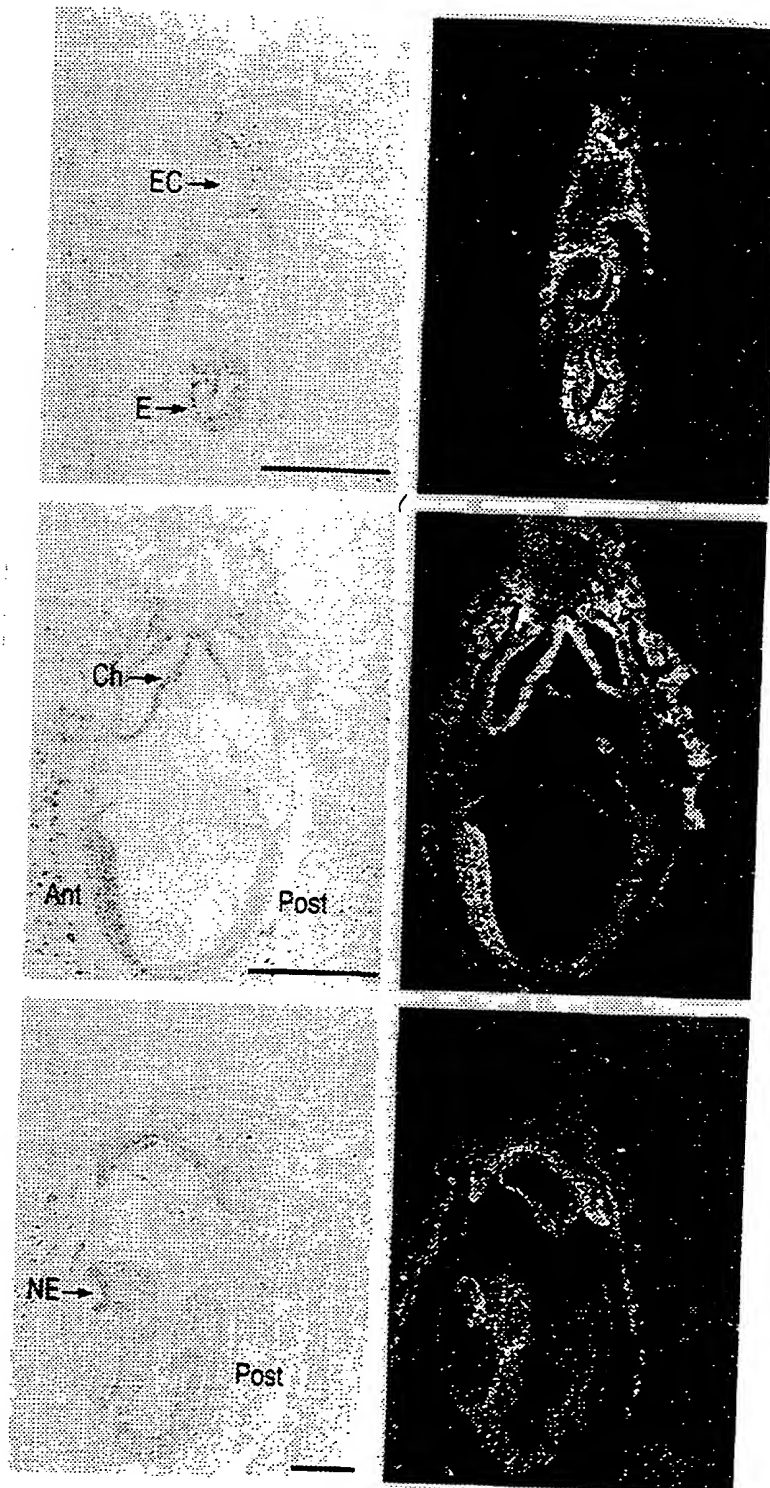


Fig. 3b

Fig. 3c



Fig. 3d

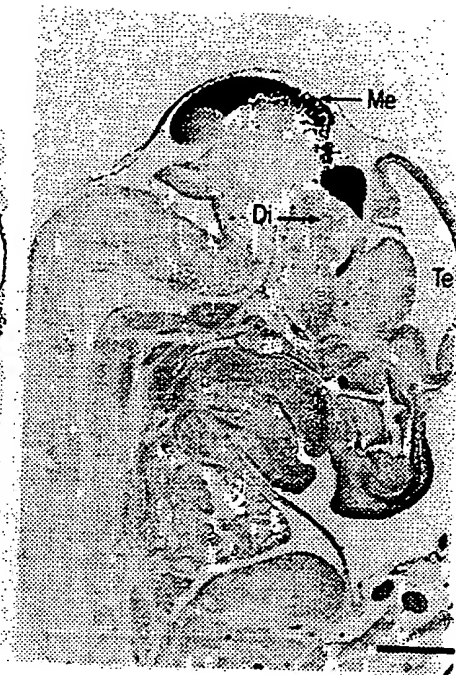
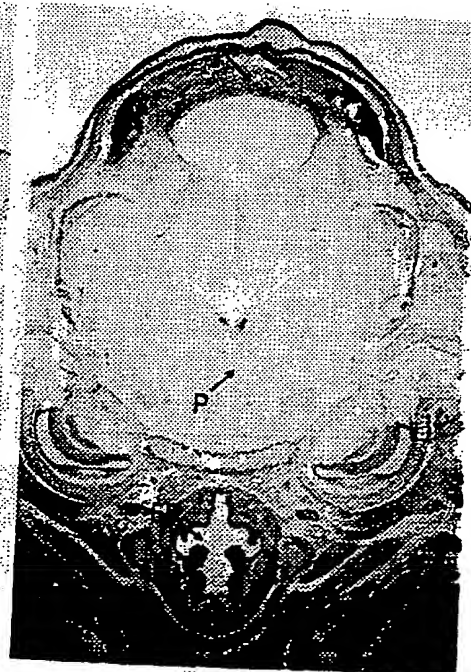


Fig. 3e



3f



3h



fig. 3g

8/18

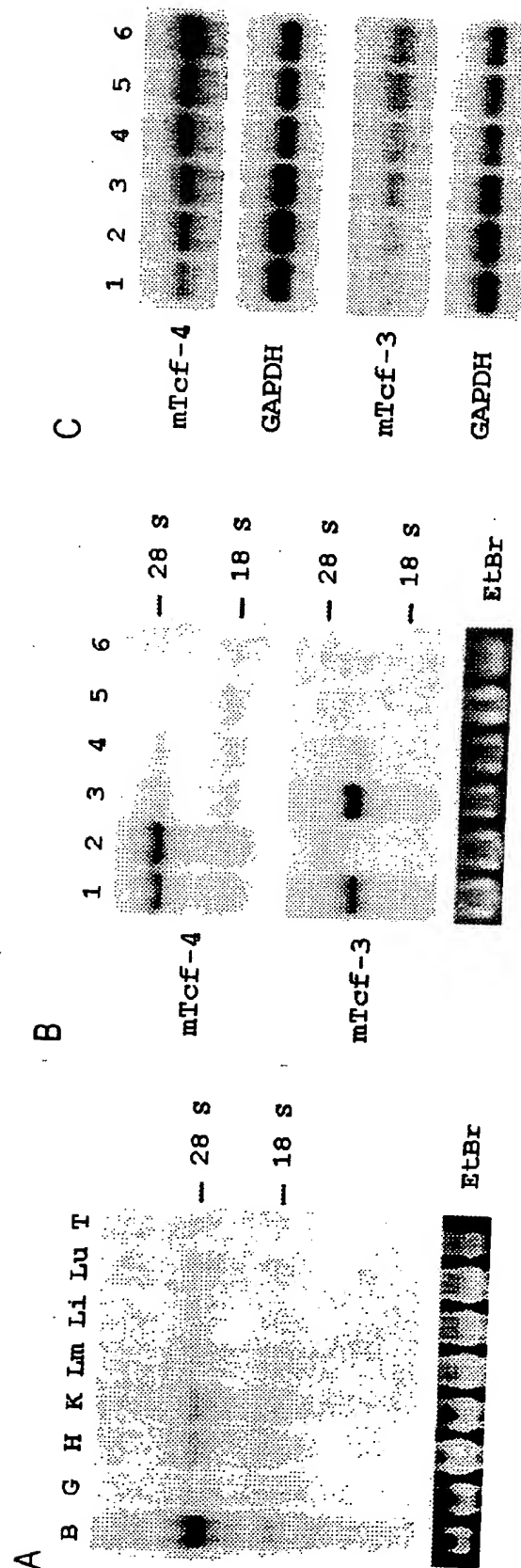


Fig. 4

9/18

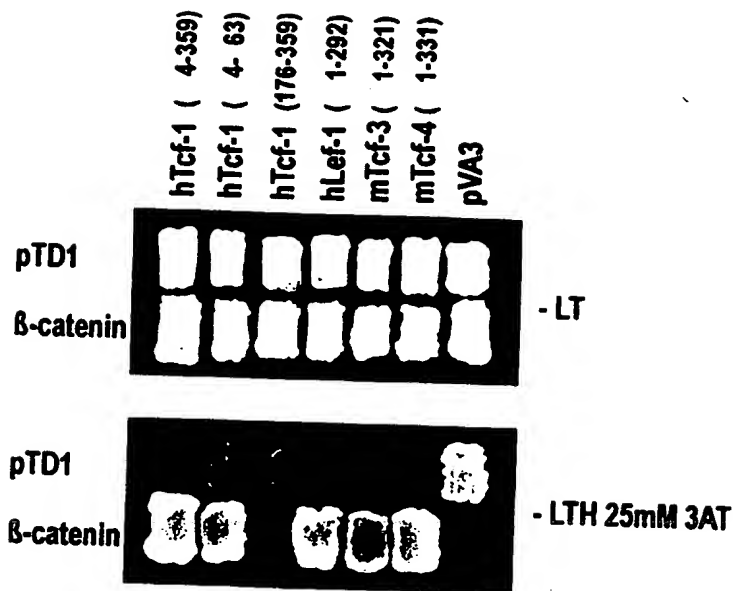


fig. 5

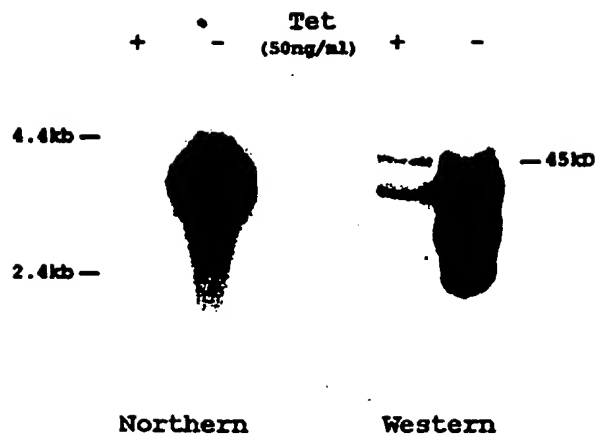
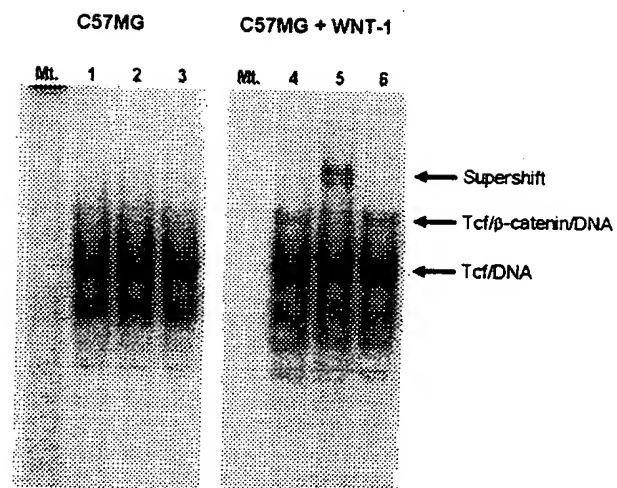
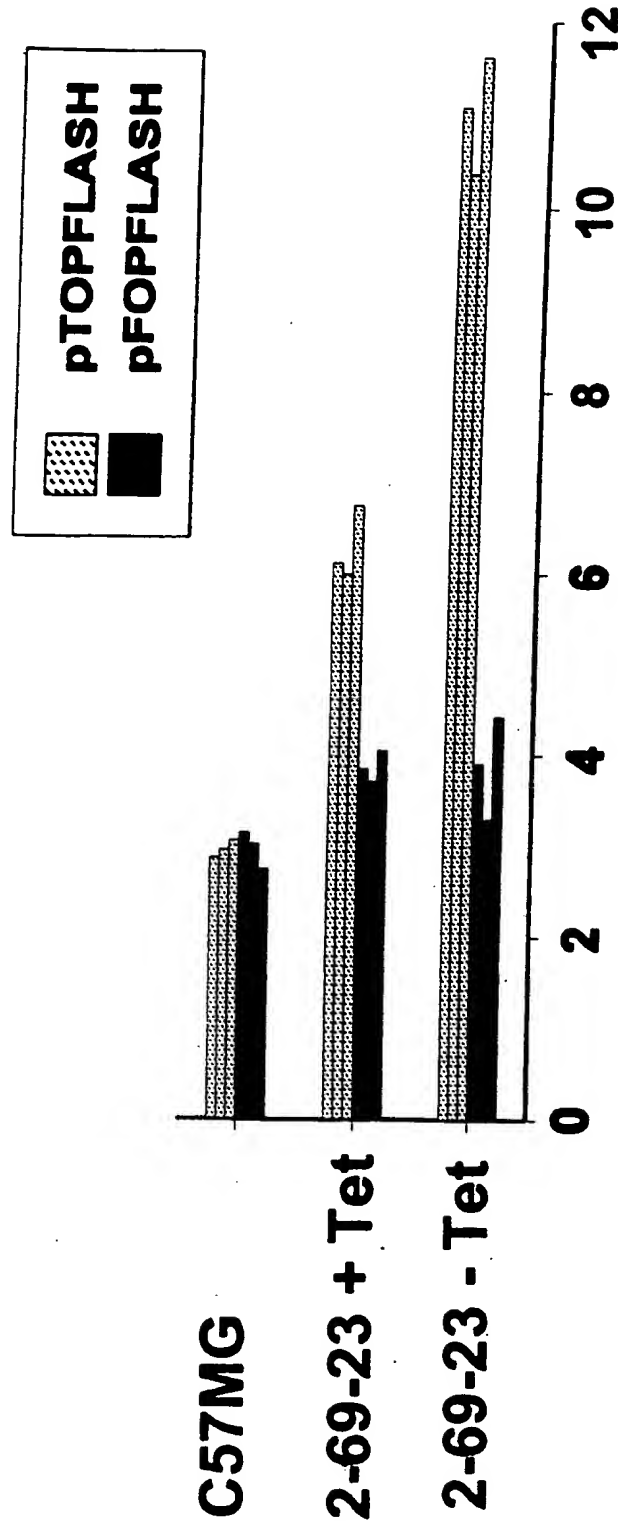


fig. 6

10/18

Fig. 7





10^3 Light units (luciferase)

Fig. 8

ATGCCC CAGCTCGGTG GTGGCCGCGG TGGCGCCGGC GGGGAGGCG GTGGCTCCGG
AGCCGGGGCA ACCAGTGGAG GGGACGACCT CGGAGCGAAC GACGAGCTGA TCCCCCTTCCA
GGACGAGGGG GGCGAAGAGC AAGAGCCGAG AAGAGCCGAG CAGCGACACG GCTTCGGCGC AGAGGGATCT
GGATGAGGTC AAGTCGTCCC TGGTCAACGA ATCGGAGAAT CAGAGCAGTA GCTCGGACTC
CGAGGCGGAG AGGCGCCCGC AGCCGCCCCG AGGATGGCGC GGACGCTTTC CAGAAGCCGC GCGACTATTT
CGCCGAAGTG AGAAGGCCCC ATGATTCCAG ACCTGAGCAG CCCGTACCTC TTTTTTTAAG GGAGGTGCGT ATCCTGGGTA
CCCCTTCCTG ATGATTCCAG ACCTGAGCAG TACAGATGAA ATGGCCCCCTC CTGATGTCC CCTGTCTCC
TGGTGGAGCG CGCACCTACC GGCATCTCC AGCACACTTG AGCACAGCAA GGCAGGCCA TGTACTCTCT
TACAGTCAAG GACACAAGGT CATCACATGC ACCCGCTGAC CGCATCTGC CCCAGAGATC GACCCAAAGA CAGGAATCCC
TCAGCACCCCT CATCACATGC TCCCCTCCCA CGCATCTGTC GTATTACCCA CTGTCTCCAG GAGCTGTCCG
CTCCCCTGCG TCCCCTCCCA CGCATCTGTC AGCTGTCACC GCTGGCTCGT CCCACAGCAA TGGCAGGCCA
CCGCCCCCTT CATCCCCTCG GCTGGCTCGT ATCCTTACCC CACACATGGT GGCTCCTGCC CATCCTGGTC
ACAAATCCCC GGTTCCTGGC AGCCGGTTCC CACCATGCTG CATTGTGAAG CAGGAGCCAG CAGCCCCCAG
CCTGAGCCCT GCAGTGAGTG CGAAATCCCC TGAATGCCCT TGCAGCCATT AACCATAATCC TGGGAAGAAA
ACCTCACGTG AAAAAGCCCC GAGTGTACCC TGAAGGAAAG CAAATACTAT GAGCTTGCCC GGAAAGAACG
GGTGGTGGCC CTGTCAAGAG AAGAACAGGC GTCAGCCCCG GACAACTATG GGAAGAAGAA
GTGGCACAAC GCAGCTTCAC GCGCAGCTCT ACCCAACCTG

Fig. 9A

GAAGAGGAAG AGAGAAAGC AGCTGTCACA GACACAGTCT CAGCAGCAAA TCCAAGAGGC
AGAGGGTGCT CTGGCCTCTA AGAGCAAGAA GCCATGCATT CAGTACCTGC CCCCTGAGAA
GCCTTGTGAT AGCCCTGCCGT CTTCCTATGG CAGCGCGCTG GACTCCCCCTG CGACGCCCTC
CGCAGCCCTTG GCTCACCAG CTGCCCCTGC AGGCCCGGC CCAGCTGGCT CTCCACTCCG CTGCCCTCCT
GTCGCTCAC ACCAAACCGG GCGCCGCCA GCAACTCCAG CCAGATGGGC AGCCAGCCCC CACTCCTATC
GTCGGCTAAG CCTTGGCT CATGCCACC AGGCCCTCCC CTGCTACAG GCCCAGCCTC TTTCCCTGGT
CAGGCCCTTG CCTTGGCT CATGCCACC AGGCCCTCCC CTGCTACAG GCCCAGCCTC TTTCCCTGGT
AGCTACGCTC CATGCCACC AGGCCCTCCC CTGCTACAG GCCCAGCCTC TTTCCCTGGT
CACCAAGTCT GCCACTAAG CAGAGGAGAG GAAGGGGAA ACACAAAGGA AGCATATTG GTCATATTT
AGTAATGATT CAGAGGAGAG GAAGGGGAA ACACAAAGGA AGCATATTG GTCATATTT
GACCACTCTG GACTGTTCTG TAAAGTGACT GGTAACAGCA GTGCCTTACC ACTGTAGATA
TAACCAGTAG CTGATCTTCA GGCTTTTTT

Fig. 9A continued

GCTGATCTCC
 GGAAAGGGAT
 CAGCTCCTCC
 TAAATCCCCG
 GCCACCGTAT
 CAACGGATCG
 TGTCCAAGCA
 AGCACACATC
 CACGCCCTCTC
 ACCAGCTGAC
 TCCATATTAC
 AGTACCACAG
 CCCCACAGCG
 CCATCACACT
 GCAGGAATCC
 AAAGGAAGAA
 TATGAAAGAG
 CAACCAGATT
 CGAGCTGGCC
 GGATAACTAT
 ACACAGCGAA
 ATGCCGCAGC
 TTCAAAGACG
 TTAGCCGATG
 GATTCCGAGG
 GAAAGTTTGG
 CCCGGCTACC
 CTCTCGCCCA
 GGAAGCCTCC
 GTTTCGAACA
 ATCACGTACA
 GTAGACCCCA
 CCGCTGTCGC
 CAAGGTCAGC
 CTGACAGTCA
 CTGCACACGA
 TCCCAGAGTG
 GAGAAGAAGA
 ATGAGAGCGA
 CTCGGGCGCA
 CGGAAGGAAC
 GGGAAGAAGA
 TGTTTCCTAA
 TGAACGGCGG
 AAGCGGAGCA
 TCAAGTCCTC
 CGAAAGACG
 AAGAAGCGGC
 CCTTCATCAT
 CCGCCCGAAC
 AGAGCAGACA
 AAGTACCGGT
 GCAATGAACA
 AAACAGGAAT
 CCGGCACCGT
 CTGTGTACCC
 ACGCATCTAT
 CCGGCATCCC
 ACGTCGGCTC
 AGCCCACAT
 AGGTGGTGGC
 GGTGGCACGC
 GACAGCTTCA
 AGAAGAGAAA
 ATCCTTGCCT
 TGAGGAGAT
 GGAGAGAAG
 GCTGCTCAAT
 GCCTCCGCCT
 CAAGAGGCAA
 GATCCCCGAC
 CTATCTTCAG
 AACCCTCAAG
 GGTGCAACAC
 CTTCAACCCG
 CCCAAGGCCT
 AGGACAAATC
 AATCACGACA
 GTCTAGGTTT
 TCACCCGGCC
 ACTCCACAGC
 AAAGAAGCCC
 CGAATGCACA
 CCTGTCCAGG
 CATGCAGCTG
 AAGAGACAAG
 TTCGCTTCCT
 GACCTAGGCG
 AACTCGGAAA
 GAATCAGAGA
 CGCTCCGAAA
 GATGGAGGGC
 CTGACGAGCC
 ATGAAATGGC
 GATGCTCGTT
 CCCCAACCATG
 GGAATCCAC
 CCGCACCTC
 CCCCATCCGC
 GGAGGATTCA
 CCTCCCCATA
 ATCGTCACAC
 TCAAAGCATC
 CTTAATGCAT
 TTGAAAGAGA
 GAAGAACAGG
 TACCCTGGCT
 CAGCCGGGGG
 CCGATCACAG
 ACCTGAGCGC
 CTAACGACGA
 ACTCCTCGGC
 CGAATCAAGA
 GTTCCGAGA
 TCTTTAAGGG
 CCTACCTCCC
 CACTGCTTGA
 CGCCGTCGCC
 TCCACCCACT
 CTCCGCACCT
 CAGATATCTC
 TAGGATGGTT
 GACATCCCTA
 TGGTCCCTCC
 CGACAGTCAA
 AGGACTCCAA
 TCATGTTGTA
 GTGCAGCCAT
 CAAAATACTA
 GGTCTGCACG
 AAACCAACGA
 ACCTGAGCGC

Fig. 9B

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TCCTAAGAAA TGCCGAGCGC GCTTTGGCCT TGATCAACAG AATAACTGGT GCGGCCCTG
CAGTCTTTGA ATTTGGAATA TTATGATGGA GAAAAAATAA AGTGC GTTCG CTACATACAA
GGTGAAGGCA GCTGCCTCAG CCCACCTCT TCAGATCCAA GCTTACTAGA CTCGCCCTCCC
CCCTCACCGC ATCTGCTAGG CTCCCCTCCC CAAGACGCCA AGTCACAGAC TGAGCAGACC
CAGCCGCTCT CGCTGTCCCT GAAGCCTGAT CCTCTGGCC ACCTGTCCAT GATGCCCTCCG
CCACCTGCGC TCCTGTGGC CGAAGCTGCC CACGGCAAGG CCTCTGCCCT CTGTCCCAAT
GGGCTCTGG ACCTGCCACC TGCCGCTCTG CAGCCGTCCA TGGTCCCTTC CTCATCGCTC
GCACAACCAT CAACTTCTTC GTCTTTAGAA TAGCTCCGCC TGGCTGGAAC GCAACCCAG
CCTCTGTCTC TGGTGACCAG GTCTTTAGAA TAGCTCCGCC TCCTCGTCCT CCATGCCCTT
GCTTGATTGA AGTGTTTCCT TCTGGTTCTT GTGCGCCCT GGTACATGA GTTAATGTTT GAAAGCTTTT
GTTTTGTACT CTTTAATTTT TGCCCGCCGT TCTTATTCA ATTTCTCCTT TTAATAATGT AGATGAGAAA
TTGGTCAATA TTGACCCAT TCTACCAA AAATTTTATCC GAAAAATAAC AATACAGGAC TCTTTCGAGG
AACCTCATGA TTCTACCAA AAATTTTATCC GAAAAATAAC AATACAGGAC TCTTTCGAGG
CCCTGTAATT GGAATGAGTC CACTTTAAAT CCTTTAACGA GGATCCATTG GAGGGCAAGT
CTGGTGCCAG CATCCGGGT A

```

Fig. 9B continued

ATGCCGCAGC TGAACGGCGG TGGAGGGGAT GACCTAGGCG CCAACGACGA ACTGATTTC
 TTCAAAGACG AGGGCGAACA GGAGGAGAAG AGTCCGAAA ACTCCTCGGC AGAGAGGAT
 TTAGCTGATG TCAAATCGTC TCTAGTCAAT GAATCAGAAA CGAATCAAAA CAGCTCCTCC
 GATTCCGAGG CGGAAAGACG GCCTCCGCCT CAAGAGGCAA GATGGAGGC TCTTTAAGG GCCACCGTAT
 GAAAGTTGG AAGAAGCGG CCTTCATCAT GATCCCCGAC ATGAAATGGC CCACTGCTGA TGTCCAGGCA
 CCCGGCTACC CCGCCCGAAC CTATCTCCAG GATGCCCGGT CCTCACCCCT TCCACCCCTT CACGCCCTT
 GTCTCGCCCA AGAGTAGACA AGCCTCAAG GGTGCAGCAC CTTACGCGG GGAACCCAC CTCCACACTT ACCAGCCGAC
 GGGAGCCTCC AAGTGCCAGT GTTACGCGG CCCACGGCT AGGACAAATC AATCACGACA GTCCAGGTT
 ATCACGTACA GCAATGAACA CTTACGCGG CCCACGGCT AGGACAAATC AATCACGACA GTCCAGGTT
 GTAGACCCCA AACAGGAAT CTGGCACCGT AGGACAAATC AATCACGACA GTCCAGGTT
 CCACTATGCG CAGTGATACC ATGCTTCCGT GTCCAGGTT GCATCCGGC ACTCCATAGT TCAAAGCATC
 CAAGGTCAC CAGTGATACC ATGCTTCCGT GTCCAGGTT GCATCCGGC ACTCCATAGT TCAAAGCATC
 CTGACCGTCA ATGCTTCCGT GTCCAGGTT GCATCCGGC ACTCCATAGT TCAAAGCATC CTTAATGCAT
 CTACACACGA CGGGCATTC ATGCTTCCGT GTCCAGGTT GCATCCGGC ACTCCATAGT TCAAAGCATC
 TCCAGAGTG ATGTCGGCTC AGCCCCACAT AAAGAAACCT TGAGTGCACG ACTGTCCAGA GAAGAGCAAG
 GAAAGAGA AGCCCCACAT AAAGAAACCT TGAGTGCACG ACTGTCCAGA GAAGAGCAAG TACCCCGGCT
 ATGAGAGCAA AGGTGATGC GGTGGCATG GACAGCTTCA TATGCAACTG AAGGGACAAG CAGCCGGGAG
 CTGGGGCGA GGTGGCATG GACAGCTTCA TATGCAACTG AAGGGACAAG CAGCCGGGAG GCTGCCCTCAG
 CGGAAGGAGC GACAGCTTCA TATGCAACTG AAGGGACAAG CAGCCGGGAG GCTGCCCTCAG
 GGAAGAAGA AGAAGAGGAA AAGGGACAAG CAGCCGGGAG GCTGCCCTCAG
 AGTGCGTTCG CTACATACAA GGTGAAGGCA GCTGCCCTCAG
 GCTTAC

Fig. 9C

ATG CCGCAGCTGA ACGGCGGTGG AGGGGATGAC CTAGGCGCCA ACGACGAACT
 GATTTCCTTC AAAGACGAGG GCGAACAGGA GGAGAAAGAGC TCCGAAAACT CCTCGGCAGA
 GAGGATTTA GCTGATGTCA AATCGTCTCT AGTCAATGAA TCAGAAACGA ATCAAAACAG
 CTCCTCCGAT TCCGAGGCGG AAAGACGGCC TCCGCCTCGC TCCGAAAGTT TCCGAGACA
 ATCCCCGGAA AGTTTGAAG AAGCGGCCAA GAGGCAAGAT GGAGGGCTCT TTAAGGGGCC
 ACCGTATCCC GGCTACCCCT TCATCATGAT CCCGACCTG ACGAGCCCCT ACCTCCCAA
 GCGATCCGTC TCGCCACCG CCCGAACCTA TCTCCAGATG AAATGGCCAC TGCTTGATGT
 CCAGGCAGG AGCTCCAGA GTAGACAAGC CCTCAAGGAT GCCCGGTCCC CATCACCGGC
 ACACATTGTC TCTAACAAAG TGCCAGTGGT GCAGCACCTT CACCATGTCC ACCCCCTCAC
 GCCTCTTATC ACGTACAGCA ATGAACACTT CACGCCGGA AACCACCTC CACACTTACC
 AGCCGACGTA GACCCCAAA CAGGAATCCC ACGGCCTCCG CACCTCCAG ATATATCCC
 GTATTACCCA CTATCGCCTG GCACCGTAGG ACAATCCCC CATTCCGTAG GATGGTAGT^{17/18}
 ACCACAGCAA GGTCACCCAG GTACCCCAAT CACGACAGGA CATTTCAGAC ACCCTACCC
 CACAGCTCTG ACCGTCAATG CTTCCGTGTC CAGGTTCCCT CCCCATAATGG TCCCACCACA
 TCATACGCTA CACACGACGG GCATTCCGCA TCCGGCCATA GTCACACCAA CAGTCAAACA
 GGAATCGTCC CAGAGTGATG TCGGCTCACT CCATAGTTCA AAGCATCAGG ACTCCAAAAA
 GGAAGAAGAA AAGAAGAAGC CCCACATAAA GAAACCTCTT AATGCATTCA TGTGTATAT
 GAAGGAAATG AGAGCAAAGG TCGTAGCTGA GTGCACGTTG AAAGAAAGCG CGGCATCAA
 CCAGATCCTT GGGCGGAGGT GGCATGCACT GTCCAGAGAA GAGCAAGCGA AATACTACGA
 GCTGGCCCCG AAGGAGCGAC AGCTTCATAT GCAACTGTAC CCCGGCTGGT CCGCGCGGA
 TAACTATGGA AAGAAGAAGA AGAGGAAAG GGACAAGCAG CCGGAGAGA CCAATGAACA

Fig. 9D

CAGCGAATGT TTCCTAAATC CTGCGCTTTC ACTTCCTCCG ATTACAGACC TCAGCGCTCC
TAAGAAATGC CGAGCGCGCT TTGGCCTTGA TCAACAGAAAT AACTGGTGCG GCCCTTGCAG
GAGAAAAAA AAGTGCGTTC GCTACATACA AGGTGAAGGC CCCCTCCCCG AGCTGCCTCA GCCACCCCTC
TTCAGATGGA AGCTTACTAG ATTCGCCCTCC CCAGCCTCTG TCGCTGTCCC TGAAGCCCCGA
CCGAGACGCC AAGTCACAGA CTGAGCAGAC GCCACCCGCC CTCCTGCTCG CTGAGGCCAC
CCCCCTGGCC CACCTGTCCA TGATGCCCTCC GGCCCTGGAC CTGCCCCCAG CCGCTTTGCA
CCACAAGGCC TCCGCCCTCT CAATTGCACA GCCGTCGACT TCTTGGTTAC ATTCCCACAG
GCCTGCCGCC CCCTCCTCAT CCCAGCCGCT GTCGCTCGTC ACCAAGTCTT TAGAA

Fig. 9D continued

INTERNATIONAL SEARCH REPORT

Inte lional Application No
PCT/NL 99/00106

A. CLASSIFICATION F SUBJECT MATTER

IPC 6 C12N15/12 C07K14/47 C12N1/19 C12N5/10 A01K67/027
C07K16/18 G01N33/574 G01N33/50

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C07K A01K G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	KORINEK V. ET AL.: "CONSTITUTIVE TRANSCRIPTIONAL ACTIVATION BY A BETA-CATENIN-TCF COMPLEX IN APC-- COLON CARCINOMA." SCIENCE, vol. 275, March 1997, pages 1784-1787, XP002070159 * the whole document, especially Fig. 1 *	1-10, 12-20
X	TAYLOR I. C. A.: "Mouse mammary tumors express elevated levels of RNA encoding the murine homolog of SKY, a putative receptor tyrosine kinase." JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 270, no. 12, 1995, pages 6872-6880, XP002075639 see abstract --- -/--	8,10,11

☒ Further documents are listed in the continuation of box C.

☐ Patent family members are listed in annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

21 June 1999

Date of mailing of the international search report

28/06/1999

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Mandl, B

INTERNATIONAL SEARCH REPORT

Int. Appl. No.

PCT/NL 99/00106

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EMBL database entry Aa671377; accession number AA671377; 27. November 1997; Marra et al.: 'The WashU-HHMI mouse EST project. XP002075697 see abstract	1-3
X	EMBL database entry Aa795343; accession number AA795343; 11. February 1998; Marra et al.: 'The WashU-HHMI mouse EST project.' XP002075698 see abstract	1-3
A	CLEVERS H. AND VAN DE WETERING M.: "TCF/LEF factors earn their wings." TRENDS IN GENETICS, vol. 13, no. 12, December 1997, pages 485-489, XP002075640 see the whole document	1-20
P,X	EMBL database entry Af057691 accession number AF057691; 16. April 1998; Sussman D.J.: 'Partial mRNA sequence of mouse transcription factor mTCF-3.' XP002075642 see abstract	1-4, 14
P,X	KORINEK V. ET AL.: "Two members of the Tcf family implicated in Wnt/beta-catenin signaling during embryogenesis in the mouse." MOLECULAR AND CELLULAR BIOLOGY, vol. 18, no. 3, March 1998, pages 1248-1256, XP002075641 see the whole document	1-20

INTERNATIONAL SEARCH REPORT

International application No.

PCT/NL 99/00106

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Although claim 20, as far as an in vivo application is concerned, is directed to a diagnostic method practiced on the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. ☐ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.